Neurovascular responses to sequential deep inspirations assessed via laser-Doppler perfusion changes in dorsal finger skin

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ABSTRACT

Rapid and deep inspirations trigger sympathetic mediated transient vasoconstriction of skin arterioles (inspiratory gasp response, IGR). Suitability of IGR for assessing neurovascular function or acute effects of interventions depends on its reproducibility. We determined the short-term variability of IGR in 28 healthy volunteers (age 19-57, 14 male) in whom finger dorsum blood perfusion was monitored by laser-Doppler during 21 sequential IGR, each separated by two minutes. IGR was quantified as the minimum perfusion during each IGR expressed as a percentage of each immediately preceding two-minute perfusion average. Results are mean ± sem. Overall, IGR was 72.3 ± 3.3% with a slight but insignificantly higher value for males than females (77.5 ± 5.0% vs. 67.2 ± 4.0%). No tendency for adaptation or sequential change in IGR occurred over the 21 sequential trials. The coefficient of variation (CV) for single IGR ranged from 21.4% to 35.6% and averaged 29.4 ± 1.0%. If triplicate IGR were used, CV was reduced to 19.2 ± 2.7. Using all 21 responses as an estimator slightly reduced this to 18.6 ± 2.4%. For within subject variations, which affects assessments of acute interventions, the absolute percent difference between two time separated IGR samples depended on the number of IGR included in each sample and the time between initial and final samples. For our data set, a sample of six IGR was optimal and yielded an overall difference of 4% for zero delay and 14% for a 20 minute delay between initial and final samples. Results suggest that the minimal detectible differences in IGR between groups are at best limited by a CV of about 20% and sequential differences are limited to between 5-15% if a sample size of six IGR is used.
BACKGROUND

A vasomotor reflex that is triggered by a rapid and deep inspiration causes an arteriolar vasoconstriction and induces a transient decrease in skin blood flow. Reports of this phenomena appeared over 50 years ago\(^1\)-\(^2\), but many details of the sympathetic neural pathways involved are not yet known. Although the reflex causes vasoconstriction, an initial small blood flow increase may precede it and often a flow increase follows (figures 3-4).

This inspiratory gasp vascular response (IGVR) has most often been measured on plantar aspects of toes and fingers with laser-Doppler\(^3\)-\(^4\) or photoplethysmography\(^5\) and has been used to study aspects of neurovascular function in many conditions including diabetes\(^6\), Raynauds phenomena\(^7\), erythromelalgia\(^8\) and leprosy\(^9\).
An important issue related to the use and interpretation of IGVR findings is its variability within the same subject and among subjects. Factors that may affect the magnitude of the vasoconstictive component include skin temperature, age, gender and vital capacity\(^4, 10\). This variability may represent an intrinsic limit to the utility of the IGVR tests. However, systematic studies of the magnitude of variability and its implications for experimental design & interpretation of findings are sparse. Such variability affects the ability to detect possible differences when comparing normal subjects to patient groups and also affects the ability to detect changes that might be induced by rapidly acting therapeutic interventions in patients with suspected neurovascular deficits.
OBJECTIVES

The present study was undertaken to characterize key features of normal IGVR variability and to explore sampling strategies that might minimize the impact of this variability.

Specifically, it was our initial objectives to:

(1) Estimate the extent to which the number of sequential IGVR samples used to estimate mean responses for individual subjects effects variability among subjects.

(2) Estimate the extent to which IGVR sample size effects the ability to detect acute changes in IGVR responses that would be associated with acute interventions in individual subjects.
METHODS

SUBJECTS: Twenty-eight volunteers (14 male) were studied. Subjects had no history of cardiovascular or respiratory abnormality, hypertension or diabetes.

PROCEDURES: Subjects sat in a height adjustable chair with hands placed palm down on a support surface. A laser-Doppler probe was placed on the right index finger dorsum (figure 1A). A small thermocouple was placed under the probe and the finger wrapped with elastic self-adhering bandaging material (figure 1B). The hand was covered with a towel and skin temperature monitored. Testing began when a steady state was reached (15-20 minutes).

During this interval subjects were instructed as to the required breathing maneuver and were given chances to practice. The instruction was to take a deep and rapid inspiration starting at the end of a normal quiet expiration and hold it for 10 seconds (figure 2).
PROTOCOL: The test protocol consisted of a series of 21 sequential inspiratory gasps (IG) taken at two minute intervals. Average finger skin blood perfusion (SBF) during two-minute intervals preceding each successive gasp were used as the reference perfusion for its following inspiratory gasp vascular response (IGVR).

IGVR was calculated using the minimum SBF during the gasp ($SBF_{\text{min}}$) and the average SBF ($SBF_0$) as shown in figure 3. Skin and room temperatures were continuously monitored and recorded every two minutes corresponding to each of the inspiratory gasps.

After the test, blood pressure was measured and an occlusion (200 mmHg) of the brachial artery for three minutes was used to determine the laser-Doppler biological zero. This value was routinely subtracted from all raw SBF data prior to analyses.
ANALYSES

The dependence of overall variability among subjects on the number of sequential IG samples used to estimate each subject's mean IGVR was determined as shown in figure 4.

To estimate the extent to which IG sample-set size affects the ability to detect acute (within subject) changes in IG responses, sample-set sizes of 1, 2, 3, 4, 5 and 6 sequential IGVR were used as illustrated in figure 5.
Initial Setup and Probe Placement

Figure 1
Inspiratory Gasp Response

Figure 2

1600
SBF
(mv)

Note "Flomotion" Pattern

Note Arterial Pulsations

INSPIRE

HOLD

EXPIRE

10 sec
Calculation of IGVR

\[
\text{IGVR} = \frac{\text{SBF}_0 - \text{SBF}_{\text{min}}}{\text{SBF}_0} \times 100
\]

Figure 3
Example of Estimating Overall SD using Triplicate Sample-Sets

This sampling strategy results in 19 triplicate samples for each of the 28 subjects. Each sample-set provides a separate estimate of the mean IGVR for that subject. To estimate an overall SD, the SD of the estimated mean of each sample-set is averaged across subjects and the average of these 19 is used as an estimate of the overall SD of the mean IGVR for the full group. The same approach was used for samples-set sizes of 1, 2, 4, 5, 6 and 10 sequential IG samples.

Figure 4
Sampling Strategy to Estimate SD of Separated Sample-Sets

Effects of IG sample-set size on the ability to detect acute changes were estimated with set sizes of 1-6 IGVR. For each set, SD of differences between 1\textsuperscript{st} & 2\textsuperscript{nd} sets were determined. As the goal was to estimate detectible levels of change between "baseline" & an "intervention", 1\textsuperscript{st} sets simulated baselines & sets starting at 10 & 20 minutes afterwards simulated intervention responses. The diagram shows a 20" separation example. For triplicate samples, responses 1-3 are the 1st set & responses 8-10 are the response set an so on up to a sample-set size of six. Correlations between 1st & 2nd sets were also determined. Results were used to estimate a minimum number of subjects needed to detect an IGVR change of either 10 or 20% of the overall mean for $\alpha = 0.05$ and a power of 90%.

Figure 5
## Summary of Subject Data

<table>
<thead>
<tr>
<th></th>
<th>Male (N=14)</th>
<th>Female (N=14)</th>
<th>Total Group (N=28)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Age (years)</strong></td>
<td>32.9±7.9</td>
<td>33.3±13.2</td>
<td>33.1±10.7</td>
</tr>
<tr>
<td><strong>Height (cm)</strong></td>
<td>177±6.3</td>
<td>167±5.8</td>
<td>172±8.0</td>
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<tr>
<td><strong>Weight (Kg)</strong></td>
<td>82.4±11.0</td>
<td>62.2±9.0</td>
<td>72.3±14.3</td>
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<tr>
<td><strong>Pressures (mmHg)</strong></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Systolic</td>
<td>128±20</td>
<td>117±13</td>
<td>123±17</td>
</tr>
<tr>
<td>Diastolic*</td>
<td>89±13</td>
<td>79±8</td>
<td>85±12</td>
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</tbody>
</table>

Values are mean ± sd. *parameter values significantly greater for males (p<0.01).
## Estimated Minimum Number of Subjects to Detect a 20% IGVR Difference Between Groups

<table>
<thead>
<tr>
<th>IG’s in each sample set</th>
<th>Number of sequential sample sets</th>
<th>Estimated overall mean</th>
<th>Variance Across Subjects</th>
<th>Minimum Subjects per group to detect 20% difference*</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>21</td>
<td>72.2 ± 3.0</td>
<td>21.1 ± 4.4</td>
<td>46</td>
</tr>
<tr>
<td>2</td>
<td>20</td>
<td>72.1 ± 2.4</td>
<td>19.2 ± 3.2</td>
<td>38</td>
</tr>
<tr>
<td>3</td>
<td>19</td>
<td>72.0 ± 2.3</td>
<td>18.7 ± 3.0</td>
<td>36</td>
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<tr>
<td>4</td>
<td>18</td>
<td>71.9 ± 2.2</td>
<td>18.3 ± 2.7</td>
<td>34</td>
</tr>
<tr>
<td>5</td>
<td>17</td>
<td>71.8 ± 2.0</td>
<td>18.1 ± 2.5</td>
<td>34</td>
</tr>
<tr>
<td>6</td>
<td>16</td>
<td>71.7 ± 1.8</td>
<td>18.0 ± 2.2</td>
<td>33</td>
</tr>
<tr>
<td>10</td>
<td>12</td>
<td>71.4 ± 1.1</td>
<td>17.6 ± 1.4</td>
<td>32</td>
</tr>
<tr>
<td>21</td>
<td>1</td>
<td>72.2 ±</td>
<td>16.9 ± 2.3</td>
<td>29</td>
</tr>
</tbody>
</table>

*Tabulated values of N are for an $\alpha$ of 0.05 and power of 90%
Estimated Minimum Number of Subjects for Detecting 10% and 20% changes in IGVR

<table>
<thead>
<tr>
<th>IG Set Size</th>
<th>SD and Correlation of paired-differences</th>
<th>Minimum Required Subjects*</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>10 minutes</td>
<td>20 minutes</td>
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<tr>
<td></td>
<td>SD</td>
<td>R</td>
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<tr>
<td>1</td>
<td>17.1</td>
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<tr>
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<td>5</td>
<td>10.6</td>
<td>0.844</td>
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<tr>
<td>6</td>
<td>10.7</td>
<td>0.842</td>
</tr>
</tbody>
</table>

*Tabulated values of N are for an $\alpha$ of 0.05 and power of 90%

Figure 9
Paired-Difference SD

Simulated IGVR pre-post responses separated by 10 or 20 minutes

Figure 10
Paired Correlations

Simulated IGVR pre-post responses separated by 10 or 20 minutes

Correlation (r)

IG Sample-Set Size

Figure 11
Minimum N to Detect 10% & 20% Changes

N of Subjects Needed vs. IG Sample-Set Size

Sample-Set Separation
- △ = 10%
- △ = 20%

- 10 minute
- 20 minute

Figure 12
1. In this group of 28 healthy subjects the overall mean perfusion decrease induced by 21 sequential inspiratory gasps and measured at the finger dorsum was 72% of each responses immediately preceding two-minute baseline blood perfusion.

2. This magnitude of IGVR is comparable to values reported for those obtained at finger palmer surface that is rich in arterial-venous anastamoses (AVAs). This suggests that the magnitude of the IGVR is not fully dependent on AVA presence.

3. Variability of the IGVR, both within and across subjects, depended on the number of sequential sample-sets included in the estimation of the mean IGVR and on the time separation between sample-sets for within subject measures.
4. The ability to statistically detect differences in IGVR between normal subjects and patients with suspected neurovascular deficits thus depends on the number of IGVR samples used to characterize the mean response and on the number of subjects N.

5. Similarly, the ability to detect acute changes in IGVR, potentially associated with effects of rapidly acting therapeutic interventions that modify IGVR, depends on the IG sample-set size, the correlation between time separated sample-sets and on N.

6. The analyses provide a framework for, and specific estimates of, the minimum number of subjects needed to detect specified IGVR differences between groups or changes in IGVR after such interventions.

7. Application of these findings to results reported in the literature suggests that some previously drawn conclusions lack suitable statistical underpinnings.
REFERENCES

4. du Buf-Vereijken PWG et al., Skin vasomotor reflexes during inspiratory gasp: Standardization by spirometric control does not improve reproducibility. Int J Microcirc 1997;17:86-92
Additional Example Responses
Inspiratory Gasp Response

SBF (mv)

Inspire →

Expire ←

10 sec
Baseline Mean ± SD
139±11.9

Deep Breath Release

20%Δ

Slower Modulation Pattern

Cardiac Pulses

137±12.1

10 sec
$Q_{\text{avg}} =$ avg perfusion during previous 120 seconds

$\delta = 50\%$
Inspiration Effect on Heart Rate

LDF

HR

Lemagx30
Inspiration Effect on Heart Rate

![Graph showing the effect of inspiration on heart rate](image-url)
[Image 54x72 to 270x360]
[Image 54x426 to 270x714]
[Image 283x228 to 562x580]

A Laser-Doppler Probe

B Coban Wrap

C Magnetic Pad

D Disk Magnet